

# ImmunoTools *special* Award 2017



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## **DGK zeta limits cytokine-dependent expansion of CD8<sup>+</sup> T cells with broad antitumor capacity**

The immune system defends the body from foreign invaders. In cancer, tumors disguise as self-body cells and evade immune attack. For this reason, it is important to identify the mechanisms that stop T lymphocytes from recognize and destroy tumors.

The diacylglycerol kinases (DGK) negatively regulate diacylglycerol (DAG)-mediated signaling by catalyzing the conversion of DAG to phosphatidic acid. In T lymphocytes two DGK isoforms: DGK $\alpha$  and DGK $\zeta$  limit activation of the Ras-ERK pathway by metabolizing the DAG generated after TCR triggering (*Merida, 2015*). Elevation of the two isoforms is observed when T cells expressing Chimeric Antigen T cell Receptors (CAR T cells) invade solid tumors, suggesting that DGK limit antigen-dependent tumor elimination (*Riese, 2013*). In agreement, DGK $\alpha$  expression in renal tumor-infiltrating CD8<sup>+</sup> T cells correlates with impaired cytotoxic responses (*Prinz, 2012*) and DGK $\zeta$  deficient mice develop smaller tumors in antigen dependent models (*Riese, 2011*).

Currently the effectiveness of IL-2 and IL-15 immunotherapy has been proved to boost antitumor immune response. IL-2 or IL-15 cytokines drive expansion and differentiation of cytotoxic CD8<sup>+</sup> T cell subsets that express natural killer receptors and eliminate targets via antigen-independent killing. Using DGK $\alpha$  and DGK $\zeta$  deficient mice, we are investigating antitumor capacity of a CD8 T cell population with high expression of the IL-2/IL-15 common  $\beta$  chain (CD122) that respond to IL-2/ IL-15 independently of antigen recognition. DGK $\zeta$  but not DGK $\alpha$  deficient mice show an increase in this population and enhance IL-2/IL-15 responses in an antigen-independent manner. We have recently demonstrated using an aggressive BALB/c-derived B cell lymphoma (A20 cells) that DGK $\zeta$  deficient mice develop smaller subcutaneous tumors and resolve it faster than wild type mice (*Andrada, 2017*).

*Ex vivo* incubation of CD8<sup>+</sup> T cells with IL-2 or IL-15 in the absence of antigen stimulation promotes differentiation of an innate-like cytotoxic cell population. These

cells called cytokine-induced killers cells have a potent antitumor activity in mouse models and in human clinical assays. We have analyzed the function of DGK $\zeta$  in the generation of cytokine-induced killers cells. We found that DGK $\zeta$  deficient cells differentiated with IL-2 presented a more cytotoxic profile than WT cells. In sight of these data, we evaluated the in vivo anti-tumor capacity of these in the A20 model. We found that mice treated with DGK $\zeta$  deficient cytokine-induced killers cells present a more rapid regression of tumors than mice treated with control cells. Currently we are analyzing the role of DGK $\zeta$  in the immune response against other low antigenic and more aggressive syngenic tumors models. We are particularly interested in melanoma models in which IL-2 is approved for human treatment. Our preliminary results indicate that DGK $\zeta$  deficient mice present a better immune response in this model. We want to investigate how pharmacological manipulation of DGK $\zeta$  combined with cytokine treatment could be employed as cancer immunotherapy.

We are interested in testing **ImmunoTools** cytokine and chemokines to further investigate how DGK $\zeta$  controls the response of T cells to other cytokines and its role in the control of T cell tumor recruitment in response to chemokines. We want to demonstrate if DGK $\zeta$  abilities are related with its function in the control of polarized responses (*Andrada, 2016*). On the other hand, additional antibodies will allow us to exhaustively characterize the profile of tumor-infiltrating lymphocytes to assess the contribution of DGK $\zeta$  deficiency to the expression of T cell markers of potential use to delimit different T cells population.

#### **References:**

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**ImmunoTools** *special* AWARD for **Elena Andrada** includes 25 reagents

**PE** - conjugated anti-human IFN-gamma

recombinant human cytokines: rh IFN-gamma, rh IL-2, rh TGF-beta3

human ELISA-set (for one 96 plate): human IFN-gamma, human TNF-a

**FITC** - conjugated anti-mouse CD48, Erythroid cells

**APC** - conjugated anti-mouse CD44

mouse ELISA-set (for one 96 plate): mouse IL-17A, mouse TNF-a

recombinant mouse cytokines: rm IFN-gamma, rm IL-15, rm IP-10 / CXCL10

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