

# ImmunoTools *special* Award 2016



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## **Mechanisms of B and T cell activation induced by targeting of MSP1<sub>19</sub> antigen to two different subsets of dendritic cells via DEC205 and DCIR2 receptors**

There are mainly two Dendritic Cells (DCs) subsets in murine spleen, one that expresses the endocytic receptor CD205 (DEC205) and the alpha chain of CD8 receptor (DEC205<sup>+</sup>CD8 $\alpha$ <sup>+</sup>), and another that expresses the endocytic receptor DCIR2 but not CD8 $\alpha$  (DCIR2<sup>+</sup>CD8 $\alpha$ <sup>-</sup>). DEC205<sup>+</sup>CD8 $\alpha$ <sup>+</sup> DC subset is specialized in antigen cross presentation and CD8<sup>+</sup> T cells priming, while the DCIR2<sup>+</sup>CD8 $\alpha$ <sup>-</sup> DC is associated with priming of CD4<sup>+</sup>T cells (*Merad et al., 2013*).

Antigen targeting to these two DCs subsets via chimeric monoclonal antibodies (mAbs) linked to proteins derived from different pathogens is an efficient strategy to enhance the immunogenicity of such proteins (*Boscardin et al., 2006; Henriques et al., 2013*). Nevertheless, the mechanisms of B cell activation, as well as the cell-to-cell interactions induced by this strategy of vaccination, are not completely understood.

Thus, the main goal of this study is to analyse the antibody response and the cell-to-cell interactions among DCs, T and B cells induced by targeting a model antigen to the two DCs subsets. We will use chimeric  $\alpha$ DEC205 and  $\alpha$ DCIR2 mAbs fused with a 19 kDa fragment derived from the *Plasmodium vivax* merozoite surface protein 1 (MSP1<sub>19</sub>). *Plasmodium* spp. are the causative agents of malaria, a disease that affects millions of people annually.

To analyse the immune response induced after immunization of mice with  $\alpha$ DEC205-MSP1<sub>19</sub> or  $\alpha$ DCIR2-MSP1<sub>19</sub> chimeric mAbs, we will use ImmunoTools antibodies for the immunophenotyping of different cell populations. We intend to immunize the mice with each chimeric mAb and then evaluate the expansion and/or activation of DCs, monocytes/macrophages, T, B, and NK cells. For this purpose, we will use the different population and activation markers listed below. In addition, we intend to generate DCs in vitro from bone marrows using GM-CSF plus IL-4 or Flt3L/CD135.

These DCs will be incubated with both chimeric mAbs ( $\alpha$ DEC205-MSP1<sub>19</sub> or  $\alpha$ DCIR2-MSP1<sub>19</sub>), activated and then injected into naïve mice. The anti-MSP1<sub>19</sub> antibody titers, as well as the induction of T cell responses, will be accessed by intracellular cytokine staining.

With this project we intend to explore how each DC subset controls T and B cell activation.

#### References:

BOSCARDIN, S. B. *et al.* Antigen targeting to dendritic cells elicits long-lived T cell help for antibody responses. **J Exp Med**, v. 203, n. 3, p. 599-606, Mar 20, 2006.

HENRIQUES, H. R. *et al.* Targeting the Non-structural Protein 1 from Dengue Virus to a Dendritic Cell Population Confers Protective Immunity to Lethal Virus Challenge. **PLoS Negl Trop Dis**, v. 7, n. 7, p. e2330, Jul 2013.

MERAD, M. *et al.* The dendritic cell lineage: ontogeny and function of dendritic cells and their subsets in the steady state and the inflamed setting. **Annu Rev Immunol**, v. 31, p. 563-604, 2013.

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includes 25 reagents

**FITC** - conjugated anti-mouse CD3e, CD4, CD8a, CD19, CD80, CD90, CD134, NK-cells, isotype control IgG2b

**PE** - conjugated anti-mouse CD4, CD8a, CD11b, CD25, CD44, CD54, CD90, isotype control IgG2b

**APC** - conjugated anti-mouse CD3e, CD4, CD19, Gr-1, isotype control IgG2b

recombinant mouse cytokines: Flt3L/CD135, rm GM-CSF, rm IL-4

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